DEPARTMENT OF CLINICAL LABORATORY SCIENCES

SCHOOL OF HEALTH PROFESSIONS STONY BROOK UNIVERSITY STONY BROOK, NEW YORK 11794-8205

MICROBIOLOGY COMPETENCY EVALUATION FORM

STUDENT NAME:		CLINICAL AFFILIATE:
Section:	Duration:	Supervisor/Instructor:

Instructions:

- 1. The student must complete all pertinent objectives/checklist items in order to complete the rotation within each section. If the items are not applicable, indicate "n/a."
- 2. The evaluator(s) must document the student's acceptable performance in meeting the established tasks in the Laboratory Safety and Infection Control, Quality Control and Microbiology Procedures and Protocols categories by putting the date the task was performed. The evaluator(s) must document the student's acceptable performance in meeting the established tasks in the Microbiology Test Skills and Affective Skills category by checking the appropriate column (1 = none of the time, 2 = some of the time, 3 = most of the time, 4 = all of the time). NA = None Applicable, may also be used.
- 3. The instructor and student must sign this form after it has been completed at the end of the clinical rotation.
- 4. *Please note* that grades of **2 or 1** may result in remediation for the student and need to be brought to the attention of the Clinical Coordinator and the student.

LABORA	FORY SAFETY AND INFECTION CONTROL	DATE PERFORMED	EVALUATOR SIGNATURE
1. IDENT	FIES LOCATION OF SAFETY DEVICES		
А.	Fire Extinguishers		
В.	Fire Alarms		
С.	Fire Blankets		
D.	Eye Washes		
E.	Emergency Exits		
F.	Safety Showers		
G.	First Aid Kit		
H.	Incident Reports		
2. REVIEV	WS DEPARTMENT'S PROTOCOL FOR HANDLING PATIENT SPECIMENS		

QUALITY CONT	TROL	DATE PERFORMED	EVALUATOR SIGNATURE
PERFORMS APPI	ROPRIATE QC PROCEDURES ACCORDING TO PROTOCOL:		
A.	Reagents (latex agglutination for microbial antigens, microbial typing sera)		
B.	Media		
C.	Biochemicals		
D.	Automated and non-automated commercial identification systems		
E.	Antimicrobial susceptibility testing		
F.	Refrigerators		
G.	Incubators (aerobic, anaerobic, and CO ₂)		
H.	Freezers		
I.	Heating Block		
J.	Water Bath		
К.	Centrifuges		
L.	Biological Safety Hood		

MICROBIOLOGY PROCEDURES AND PROTOCOLS	DATE PERFORMED	EVALUATOR SIGNATURE
1. Identifies location of the Clinical Microbiology Procedure Manual		
2. Reviews protocol for the use of the Laboratory Information System. Demonstrates ability to		
enter, review and retrieve data.		
3. Reviews protocol for the collection and transport of clinical specimens		
4. Reviews protocol for rejection of specimens		
5. Reviews protocol for infection control in the laboratory		
6. Reviews protocol for processing and interpretation of:		
a. Blood culture specimens		
b. CSF specimens		
c. Body fluids other than CSF specimens		
d. Upper respiratory tract specimens		
e. Lower respiratory tract specimens		
f. Genital specimens		
g. Fecal specimens		
h. Urine specimens		
i. Wound specimens		
j. Tissue specimens		
k. Skin specimens		
1. Hair specimens		
m. Nail specimens		
7. Reviews protocol for the operation and/or use of automated and non-automated commercial		
microbial identification systems.		
8. Reviews protocol for the use of biochemical and serological tests in the identification of		
microbes.		
9. Reviews protocol for antimicrobial susceptibility testing		
10. Reviews protocol for febrile agglutinin test		
11. Reviews protocol for cold agglutinin test		
12. Reviews protocol for operation and maintenance of nephelometer		
13. Reviews protocol for operation and maintenance of ELISA/EIA plate reader		
14. Reviews protocol for operation and maintenance of fluorescent microscope		
COMMENTS:		

1 = None of the Time 2 = Some of the time 3 = Most of the time 4 = All of the time

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MICROBIOLOGY TEST SKILLS	1	2	3	4	EVALUATOR SIGNATURE
1. Disposes of hazardous waste according to protocol					
2. Decontaminates work area and spills according to protocol					
3. Uses gloves appropriately					
4. Uses the Biological Safety Hood according to protocol					
5. Labels all media and biochemicals legibly and accurately					
6. Organizes workload appropriately					
7. Records and enters test results accurately and in a timely manner					
8. Uses all micropipettors appropriately					
9. Labels tubes/slides/plates legibly and accurately					
10. Prepares dilutions according to protocol					

MICROBIOLOGY TEST SKILLS (CONT'D.)	1	2	3	4	EVALUATOR SIGNATURE
Bacteriology - Minimum # of Procedures = 10					
Stains and Biochemicals:					
1. Performs and interprets the Gram stain according to protocol					
2. Performs and interprets the catalase test according to protocol					
3. Performs and interprets the slide and tube coagulase test according to protocol					
4. Performs and interprets the oxidase test according to protocol					
5. Performs and interprets the wet mount according to protocol					
6. Performs and interprets the urease test according to protocol					
7. Performs and interprets the motility test according to protocol					
8. Performs and interprets the spore stain according to protocol					
9. Performs and interprets the Dnase test according to protocol					
Serological:					
1. Performs streptococcal grouping according to protocol					
2. Performs serotyping of <u>Salmonella</u> and <u>Shigella</u> according to protocol					
3. Performs serotyping of <u>Neiserria menigitidis</u> according to protocol					
4. Performs serotpying of <u>Haemophilus influenzae</u> according to protocol					
5. Performs bacterial antigen test					
6. Performs cryptococcal antigen test (usually performed in Bacteriology) according					
to protocol					
Identification of the Principle Bacteria Encountered in Clinical Laboratory					
Using identification protocol, including both automated and non-automated					
techniques, identify the following groups of bacteria:					
1. <u>Staphylococcus spp.</u>					
2. <u>Streptococcus</u> spp.					
3. Enterobacteriaceae					
4. Gram negative fermentating rods other than Enterobacteriaceae					
5. Non-fermenting Gram negative bacilli					
6. <u>Vibrionaceae</u>					
7. <u>Campylobacter</u> spp.					
8. MRSA					
9. VRE					
10. <u>Neisseria</u> spp.					
11. <u>Haemophilus</u> spp.					
12. Gardnerella vaginalis					
13. <u>Clostridium</u> spp.					
14. <u>Bacteroides</u> spp.					
15. Prevotella spp.					
16. Porphysomonas spp.					
17. Fusobacterium spp.					
18. <u>Peptococcus niger</u>					
19. Peptostreptococcus spp.		ſ			
Antimicrobial Susceptibility Testing:					
1. Using the Bauer-Kirby disc diffusion technique, perform, read and interpret					
antimicrobial susceptibility tests on aerobic and facultative anaerobic bacteria					
commonly isolated in the clinical laboratory					
2. Performs, reads and interprets MICs on aerobic and facultative anaerobic bacteria					
commonly isolated in the clinical laboratory					

MICROBIOLOGY TEST SKILLS (CONT'D.)	1	2	3	4	EVALUATOR SIGNATURE
Mycology - Minimum # of Procedures = 5					
Stains and Biochemicals					
1. Performs and reads the lactophenol cotton blue stain according to protocol					
2. Performs and reads the 10% KOH preparation with and without calcofluor white					
according to protocol					
3. Performs and reads the india ink preparation according to protocol					
4. Performs and reads the modified Kinyoun acid-fast stain according to protocol					
5. Performs and interprets the germ tube test according to protocol					
6. Performs and interprets the Trichophyton agars according to protocol					
7. Inoculates and interprets the Christensen's urea agar test according to protocol					
8. Performs, reads and interprets the casein hydrolysis test according to protocol					
9. Performs, reads and interprets the tyrosine hyrdolysis test according to protocol					
10. Performs, reads and interprets the exanthine hydrolysis test according to protocol					
11. Performs, reads and interprets the starch hydrolysis test according to protocol					
Techniques					
1. Performs, reads and interprets the Dalmau technique according to protocol					
2. Performs, reads and interprets the teased preparation according to protocol					
3. Performs, reads and interprets the scotch tape preparation according to protocol					
4. Performs, reads and interprets the slide culture according to protocol					
5. Performs, reads and interprets the hair perforation test according to protocol					
Identification of the commonly isolated Yeasts in the Clinical Laboratory					
Using identification protocol, including both automated and non-automated techniques,					
identify the following yeasts:					
1. Candida albicans					
2. Candida tropicalis					
3. <u>Candida guilliermondii</u>					
4. <u>Candida parapsilosis</u>					
5. Candida kefyr					
6. Candida krusei					
7. Toroulopsis glabrata					
8. <u>Trichosporon</u> spp.					
9. <u>Cryptococcus neoformans</u>					
10. <u>Saccharomyces</u> spp.					
11. <u>Geotrichum</u> spp.					
Identification of the Commonly Isolated Filamentous Fungi (Molds) in the Clinical					
Laboratory					
Using identification protocol, identify the following molds:					
1. Aspergillus (to include A. fumigatus)					
2. <u>Rhizopus</u> (6 Interest <u>Interest (6</u> Interest)					
3. Mucor	1	1		1	
4. Absidia	1	1			
5. Cunnighamella					
6. Syncephalastrum					
7. Aureobasidium	1				
8. Penicillium	1				
9. <u>Paecilomyces</u>					
9. <u>Paceholityces</u> 10. <u>Scopulariopsis</u>					
11. Acremonium					
12.Fusarium	+				
13. <u>Philalophora</u>	+				
15. <u>Philaiophora</u>	1	<u> </u>	I	1	

MICROBIOLOGY TEST SKILLS (CONT'D.)	1	2	3	4	EVALUATOR SIGNATURE
Mycology - Minimum # of Procedures = 5					SIGNATORE
14. Fonsecaea					
15. <u>Cladosporium</u>					
16. Wangiella					
17. Exophiala					
18. Pseudallescheria					
19. Microsporum spp.					
20. <u>Trichophyton</u> spp.					
21. Epidermophyton flocossum					
22. Sporothrix schenckii					
Identification of the Commonly Isolated Aerobic Actinomycetes in the Clinical					
Laboratory					
1. <u>Nocardia asteroides</u>					
2. Nocardia brasiliensis					
3. Nocardia otitidiscaviarum					
4. <u>Streptomyces anulatus (griseus)</u>					
(<u>Support () · · · · · · · · · · · · · · · · · ·</u>					
Parasitology - Minimum # of Procedures = 5					
Techniques					
1. Makes wet mount preparations of fecal samples according to protocol					
2. Concentrates fecal samples by the formalin ethyl-acetate concentration procedure					
according to protocol					
3. Concentrates fecal samples by the zinc sulfate flotation method according to protocol					
4. Prepares polyvinyl alcohol (PVA) fixed fecal smears according to protocol					
5. Makes wet mount preparations from the string obtained from the "string test"					
according to protocol					
6. Prepares thin blood smears according to protocol					
7. Prepares thick blood smears according to protocol					
Stains					
1. Performs and reads wet mounts of fecal samples stained with D'Antoni's or Doebell					
and Occonor's iodine solution according to protocol					
2. Performs and reads trichrome stained PVA fixed fecal smears according to protocol					
3. Performs and reads the modified trichrome stained (for microsporidia) fecal smears according to protocol					
4. Performs and reads the Kinyoun carbolfuchsin acid-fast stained fecal smears					
according to protocol					
5. Performs and reads the rhodamine-auramine O stained fecal smears according to					
protocol					
6. Performs and reads Giemsa stained thin and thick blood smears according to					
protocol					
Identify the following more commonly seen Parasites using Iodine Stained Wet Mount					
Preparations, Trichrome Stained Slides and Acid-Fast Stained Slides:					
A. Intestinal Parasites:					
Amebae:					
Entamoeba histolytica					
Entamoeba hartmanii					
	1	1	1	1	
Entamoeba coli Endolimax nana		-	-		

Danasitalagy (Contid)	1	2	3	4	EVALUATOR
Parasitology (Cont'd)		2	3	4	SIGNATURE
A. Intestinal Parasites:					
Iodamoeba butschlii					
Blastocystis hominus					
Flagellates:					
Dientamoeba fragalis					
Giardia duodenalis (lamblia)					
Chilomastix mesnili					
Ciliates:					
Balantidium coli					
Microsporidia					
Coccidia:					
Isospora belli					
Cryptosporidium <u>spp.</u>					
Blood and tissue protozoa:					
Plasmodium:					
vivax					
malariae					
falciparum					
ovale					
Babesia spp.					
<u>Trypanosoma</u> spp. (african)					
Trypansoma cruzi					
Toxoplasma gondii					
Leishmania spp.					
Intestinal nematodes:					
Enterobius vermicularis					
Ascaris lumbricoides					
Hookworm					
Trichuris trichiura					
Strongyloides stercoralis					
Cestodes:					
Taenia saginata					
Hymenolepsis nana					
Hymenolepsis diminuta					
Diphylobothrium latum					
Dipylidium caninum					
Trematodes:					
Clonorchis sinensis					
Fasciola hepatica					
Paragonimus weestermanii					
Fasciolepsis buski					
Schistosoma mansoni					
Schistosoma haematobium					
Filarial nematodes:					
Wuchereria bancrofti					
Brugia malavi			<u> </u>		
Loa loa					
Mansonella <u>ozzardi</u>					
Mansonella streptocerca					
mansonena sueptotetea	1		1	1	

= None of the Time	2 = Some of the time	3 = Most of the time	4 = All of the time

Parasitology (Cont'd)	1	2	3	4	EVALUATOR SIGNATURE
Mansonella perstans					
Onchocerca volvulus					
Ectoparasites:					
Pediculus humanus var. corporis					
Pediculus humanus var. capitis					
Phthirus pubis					
Dermacentor spp.					
<u>Ixodes</u> spp.					
Immunology/Serology - Minimum # of Procedures = 5					
1. Performs test, including all appropriate quality control, according to protocol and					
correctly records and interprets the results for:					
a. non-treponemal antibody tests (e.g. RPR, VDRL)					
b. treponemal antibody tests (FTA-ABS, MHA-TP)					
c. heterophile antibodies (e.g. Mono-Diff, Mono-Spot, Mono-Test)					
d. antinuclear antibodies					
e. streptococcal antibodies					
f. rheumatoid factor					
g. C-reactive protein					
h. Lyme antibodies					
i. Viral antibodies/antigens:					
1. Epstein-Barr					
2. Cytomegalovirus					
3. Rubella					
4. Hepatitis A					
5. Hepatitis B			1		
6. Hepatitis C			1		
7. HIV-1			1		
COMMENTS:	1				1

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NOTE: PLEASE RECORD ANY LATENESS AND/OR ABSENCES ON THE ATTENDANCE SHEET

NOTE: MID-ROTATION PERFORMANCE

A mid-rotation performance evaluation has been included in order to allow the student to recognize and improve on laboratory tests before the student's rotation is finished. Please indicate below, and inform the student of any area he/she may need to improve.

LABORATORY TEST(S) NEEDING IMPROVEMENT: (Please indicate date of comment and add your initials to any comments)

LABORATORY TEST(S) - PROGRESS REPORT: (Please indicate date of comment)

Signature of Person Completing Form:	Title:	Date:

Student Signature:	_ Date:
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